

The microbiological hydroxylation of 3 α ,17 β - and 3 β ,17 α -dihydroxy-5 α -androstanes by *Cephalosporium aphidicola*[†]

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Both 3 α ,17 β - and 3 β ,17 α -dihydroxy-5 α -androstanes are hydroxylated at C-6 β by the fungus, *Cephalosporium aphidicola* but the 3 α ,17 β -diol is also hydroxylated at C-7 α whereas the 3 β ,17 α -diol is hydroxylated at C-11 β .

Keywords: microbiological hydroxylation, steroids, *Cephalosporium aphidicola*

Models for the microbiological hydroxylation of steroids envisage three active centres on the hydroxylase, two of which bind the substrate and one delivers the oxygen.^{1,2} It has been suggested that³ if a steroid was bound to the microbial hydroxylase at C-3 and C-17, rotation about the C-3:C-17 axis could bring different sites into juxtaposition with the hydroxylating centre of the enzyme. As part of our studies⁴ on the application of the fungus, *Cephalosporium aphidicola* to microbiological hydroxylation, we have compared the hydroxylation of 3 α ,17 β -dihydroxy-5 α -androstanone **1**⁵ with 3 β ,17 α -dihydroxy-5 α -androstanone **4**⁶ in which the opposite relative configurations of the 3- and 17-alcohols might favour different orientations about the C-3:C-17 axis.

3 α ,17 β -Dihydroxy-5 α -androstanone **1** was obtained by reduction of 17 β -acetoxy-5 α -androstan-3-one to the 3 β -alcohol and inversion of the configuration at C-3 using a Mitsunobu reaction with chloroacetic acid as the nucleophile. 3 β ,17 α -Dihydroxy-5 α -androstanone was one of the products of hydroboration of 3 β -hydroxy-5 α -androstan-16-ene.⁷

Incubation of the 3 α ,17 β -dihydroxy-5 α -androstanone **1** with *C.aphidicola* gave two metabolites, **2** and **3** which were separated by chromatography. The ¹³C NMR spectrum (see Table 1) of the major metabolite **2** contained a new CH(OH) signal at δ_C 71.7. The signals assigned to C-5 and C-7 had moved downfield whilst there were γ -carbon upfield shifts for C-4 and C-8. Hence the new hydroxyl group was at C-6. The new CH(OH) ¹H NMR signal (δ_H 3.70) was a broad singlet and the 19-H signal showed a significant downfield shift consistent with a 6 β -stereochemistry for the alcohol. The ¹³C NMR spectrum of the second metabolite also revealed a new CH(OH) resonance (δ_C 67.5 ppm). There were downfield shifts for C-6 and C-8 and γ -gauche upfield shifts for C-5, C-9 and C-14 consistent with the location of the new hydroxyl group at C-7. The ¹H NMR signal (δ_H 3.83) was a broad singlet typical of a C-7 α (axial) alcohol. The metabolites were therefore assigned the structures of 3 α ,6 β ,17 β - and 3 α ,7 α ,17 β -trihydroxy-5 α -androstanes, **2** and **3**.⁸ Incubation of 3 β ,17 α -dihydroxy-5 α -androstanone **4** gave two metabolites **5** and **6**. In the ¹³C NMR spectrum of **5** the high-field resonance associated with C-11 had been replaced by a CH(OH) signal (δ_C 68.2) whilst there were downfield shifts for the signals assigned to C-9 and C-12 and a γ -gauche shift for C-8 and C-13. The new CH(OH) resonance (δ_H 4.08) was a broad singlet whilst both the 18-H and 19-H resonances showed significant downfield shifts consistent with interactions with an 11 β -hydroxyl group. The major metabolite was the 3 β ,6 β ,17 β -triole **6**. There was a new

Table 1 ¹³C NMR data for the metabolites

Carbon atom	Compound					
	1	2 ^a	3	4 ^b	5 ^b	6 ^b
1	33.0	33.9	32.0	3a.9	37.4	40.9
2	29.9	29.6	2a.9	33.a	33.2	34.0
3	65.6	66.2	66.4	71.9	70.8	70.6
4	36.9	32.8	35.5	40.6	39.2	34.7
5	39.6	41.7	31.6	46.4	46.4	50.0
6	29.1	71.7	36.3	30.5	29.1	72.8
7	32.2	39.0	67.5	34.2	32.7	43.1
8	35.9	30.3	39.6	37.2	32.3	33.0
9	55.0	54.3	46.1	55.0	58.9	56.5
10	36.6	36.0	36.3	35.1	36.5	37.5
11	20.9	20.1	20.2	22.5	68.2	22.8
12	37.6	36.5	36.3	34.3	41.8	38.8
13	43.6	42.9	42.9	48.0	45.4	47.2
14	51.5	50.7	45.3	50.4	51.1	50.1
15	23.8	23.2	22.8	26.4	25.4	26.9
16	31.0	29.9	30.4	33.6	33.7	34.3
17	81.4	81.5	81.8	80.5	79.5	79.5
18	11.6	11.0	10.1	13.8	15.7	18.9
19	12.0	14.6	10.9	18.8	20.2	19.2

^aIn CDCl₃:CD₃OD; ^bIn C₅D₅N.

CH(OH) resonance at δ_C 72.8. The signals assigned to C-5 and C-7 showed downfield shifts whilst C-4 and C-8 showed a characteristic γ -gauche upfield shift. The multiplicity of the new CH(OH) resonance was obscured by the 3-H and 17-H signals but the 19-H signal showed a downfield shift consistent with a diaxial interaction with a 6 β -hydroxyl group.

The hydroxylation of **1** at C-11 β and of **4** at C-7 α are interesting because these two centres are related by rotation about the C-3:C-17 axis. This suggests that a model involving a common hydroxylase with different substrate binding modes may be used to rationalise the hydroxylations by *Cephalosporium aphidicola*.

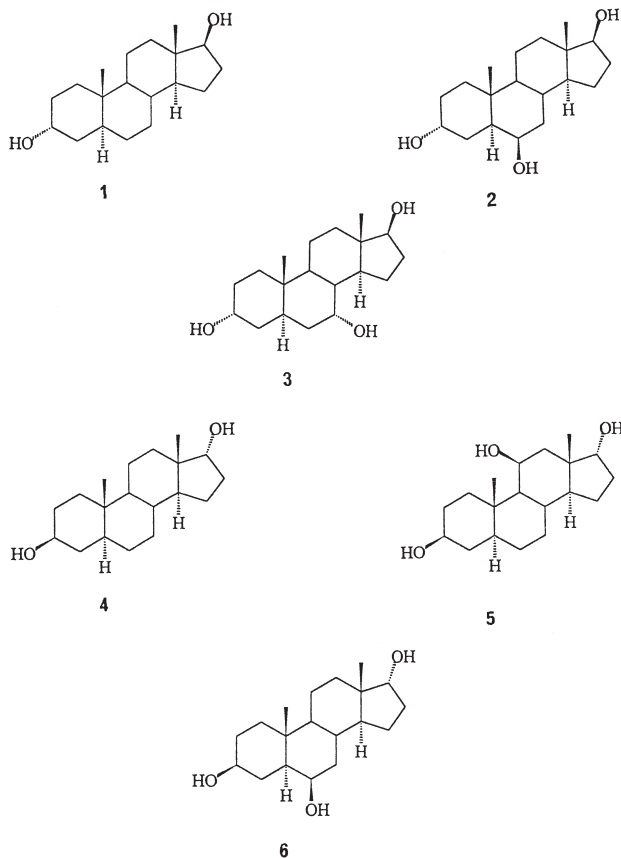
Experimental

Silica for chromatography was Merck 9385. ¹H and ¹³C NMR spectra were determined in deuteriochloroform, unless otherwise stated, at 360 and 90 MHz respectively. IR spectra were determined as nujol mulls. Light petroleum refers to the fraction b.p. 60–80°C. Extracts were dried over sodium sulfate.

Preparation of 3 α ,17 β -dihydroxy-5 α -androstanone. Diethyl azodicarboxylate (3 cm³) was added to a solution of 17 β -acetoxy-5 α -androstan-3 β -ol⁹ (3 g), triphenylphosphine (4.5 g) and dry chloroacetic acid (2.1 g) in dry toluene (90 cm³). The resulting orange solution was stirred at room temperature overnight. The solvent was evaporated and the residue was chromatographed on silica. Elution with 3% ethyl acetate:light petroleum gave 17 β -acetoxy-3 α -chloroacetoxy-5 α -androstanone (2.83 g) which crystallised from ethyl acetate as needles, m.p. 120°C, (Found: C, 67.3; H, 8.6. C₂₃H₃₅ClO₄

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[†] This is a Short Paper, there is therefore no corresponding material in *J. Chem. Research (M)*.



requires C, 67.2; H, 8.6%), $\nu_{\max}/\text{cm}^{-1}$ 1751, 1726; δ_{H} 0.78(3H, s, 18-H), 0.81(3H, s, 19-H), 0.70-1.80 (22H, overlapping multiplets), 2.03(3H, s, OAc), 4.07 (2H, s, OCCH_2Cl), 4.58(1H, t, J 8.5 Hz, 17-H), 5.11(1H, br. s, 3-H).

The chloroacetate (2.7 g) in methanol (120 cm^3) was treated with a solution of potassium carbonate (8 g) in water (40 cm^3) at room temperature for 24 h. Acetic acid (15 cm^3) was added and the mixture was stirred for 5 min. The methanol was evaporated and the residue was extracted with ethyl acetate. The extract was washed with aqueous sodium hydrogen carbonate, water, brine and dried. The solvent was evaporated to give 3 α ,17 β -dihydroxy-5 α -androstane 1 (1.8 g) which crystallised from ethyl acetate as needles, m.p. 223–224°C, (lit.,⁵ 222–224°C), $\nu_{\max}/\text{cm}^{-1}$ 3418, 3386; δ_{H} 0.73(3H, s, 18-H), 0.79(3H, s, 19-H), 0.80–1.95(22H, overlapping multiplets), 3.64(1H, t, J 8.7 Hz, 17-H), 4.05(1H, t, J 2.8 Hz, 3-H).

Incubation of substrates: The fungus, *Cephalosporium aphidicola* was grown on shake culture in 250 cm^3 conical flasks on a medium (100 cm^3) comprising (per litre), glucose (50 g), potassium dihydro-

gen phosphate (5 g), magnesium sulfate (2 g), glycine (2 g), potassium chloride (1 g) and a trace elements solution (2 cm^3). The trace elements solution contained (per litre), zinc sulfate (1.6 g), ferrous sulfate (1 g), cobalt nitrate (1 g), ammonium molybdate (1 g), copper sulfate (0.1 g) and manganese sulfate (0.1 g). The fungus was grown for 2 days before the substrate (1.5 g) in ethanol (50 cm^3) was distributed over 50 flasks. The fermentation was then continued for a further 7 days. The broth was filtered and the mycelium was washed with ethyl acetate. The broth was acidified with dil. hydrochloric acid and extracted with ethyl acetate. The extracts were washed with water and dried. The solvent was evaporated and the residue was chromatographed on silica and eluted with an increasing gradient of ethyl acetate in light petroleum.

3 α ,17 β -Dihydroxy-5 α -androstane 1 gave 3 α ,6 β ,17 β -trihydroxy-5 α -androstane 2 (101 mg) which crystallised as needles, m.p. 219°C, (Found: C, 72.1; H, 10.2. $\text{C}_{19}\text{H}_{32}\text{O}_3 \cdot 0.5\text{H}_2\text{O}$ requires C, C, 71.8; H, 10.5%), $\nu_{\max}/\text{cm}^{-1}$ 3353 (br), δ_{H} 0.77(3H, s, 18-H), 1.02(3H, s, 19-H), 0.80-1.90(20H, overlapping multiplets), 3.65(1H, t, J 9 Hz, 17-H), 3.70(1H, br. s, 6-H), 4.18(1H, m, 3-H). Further elution gave 3 α ,7 α ,17 β -trihydroxy-5 α -androstane 3 (57 mg) which crystallised as needles, m.p. 194–196°C, (lit.,⁸ 195–197°C), $\nu_{\max}/\text{cm}^{-1}$ 3450; δ_{H} 0.74(3H, s, 18-H), 0.79(3H, s, 19-H), 0.80-1.90(20H, overlapping multiplets), 3.69(1H, t, J 9 Hz, 17-H), 3.83(1H, br. s, 7-H), 4.18(1H, m, 3-H).

3 β ,17 α -Dihydroxy-5 α -androstane 4 gave 3 β ,11 β ,17 α -trihydroxy-5 α -androstane 5 (118 mg) which crystallised from acetone as needles, m.p. 221–223°C, (Found: C, 73.7; H, 10.5. $\text{C}_{19}\text{H}_{32}\text{O}_3$ requires C, 74.0; H, 10.5%), $\nu_{\max}/\text{cm}^{-1}$ 3495, 3412, 3343; δ_{H} (pyridine- d_5) 1.22(3H, s, 18-H), 1.39(3H, s, 19-H), 0.90–2.20(20H, overlapping multiplets), 3.97 (2H, m, 3-H and 17-H), 4.08(1H, br. s, 11-H). Further elution gave 3 β ,6 β ,17 α -trihydroxy-5 α -androstane 6 (203 mg) which crystallised from ethyl acetate as needles, m.p. 204–205°C, (Found: C, 73.7; H, 10.3. $\text{C}_{19}\text{H}_{32}\text{O}_3$ requires C, 74.0; H, 10.5%), $\nu_{\max}/\text{cm}^{-1}$ 3370; 32 δ_{H} (pyridine), 0.71(3H, s, 18-H), 1.39(3H, s, 19-H), 0.90–2.20(20H, overlapping multiplets), 4.0(3H, br. m. 3-, 6- and 17-H).

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